



***Plasmodium falciparum* Glucose 6-Phosphate Dehydrogenase-6-
Phosphogluconolactonase. Characterisation of Redox-Related
Networks as Contribution to the Development of Novel
Intervention Strategies**

by

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from

Mombasa, Kenya

**A thesis submitted to the Faculty of Biology and Chemistry
(FB 08) in partial fulfilment for the requirements of the
Doctor of Science Degree of Justus-Liebig-University
Giessen, Germany**

September 2008

Dedication

This thesis is dedicated to my daughter Lynn Ndimu Mwongela, my son Liam Musyimi Jnr IIIrd Mwongela, my wife Grace C. Mwangome and the entire Musyimi family.

You all inspire my life to live the dream.

Berichte aus der Biochemie

Boniface Mwongela Mailu

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D 26 (Diss. Universität Giessen)

Gedruckt mit Unterstützung des Deutschen Akademischen Austauschdienstes

Shaker Verlag
Aachen 2009

Bibliographic information published by the Deutsche Nationalbibliothek

The Deutsche Nationalbibliothek lists this publication in the Deutsche Nationalbibliografie; detailed bibliographic data are available in the Internet at <http://dnb.d-nb.de>.

Zugl.: Giessen, Univ., Diss., 2008

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Printed in Germany.

ISBN 978-3-8322-8134-2

ISSN 1434-5536

Shaker Verlag GmbH • P.O. BOX 101818 • D-52018 Aachen

Phone: 0049/2407/9596-0 • Telefax: 0049/2407/9596-9

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I Have a Dream.....

(Martin Luther King Jnr, August 28th, 1963)

The thesis was presented to the faculty of Biology and Chemistry of the Justus-Liebig University Giessen, Germany for examination on the 11th September 2008 and the thesis defence was on the 5th November 2008.

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Acknowledgements

Many individuals have contributed to the overall research achievements reported herein. My sincere gratitude goes to my supervisors, Prof. Dr. med Katja Becker and Prof. Dr. Gabrielle Klug. I would like to especially thank Prof. Katja Becker who apart from her guidance and hosting me in her research group, shared with me her wide experience in redox metabolism of the malaria parasite *Plasmodium falciparum*. I extend my heartfelt and sincere thanks for her excellent supervision, fruitful ideas, valuable advice and encouragement during the period of my study.

I am greatly indebted to all distinguished members of Prof Becker's research group both past and present members (2005-2008), who assisted me with invaluable counsel in the course of this study. Specifically many thanks go to Dr. Stefan Rahlfs who was my officemate and collaborator in all the studies, Dr. Rimma Iozef (once a chemist like me) who cloned the human 6-PGL and was a great source of inspiration and encouragement, Marina Fischer for the hands on experience on enzyme assays, Elisabeth Fischer for the great experience in *P. falciparum* cell culture, Beate Hecker thanks for your help and experience in IDO assays and gel filtration, Michaela Stumpf with the help of your magic hands we have *Pf*GluPho crystals, Kathleen Zocher thanks for your protein modelling experience, Ulrike B Z for the help with graphics, Sabine Ettinger and Santhosh Maddila for their wonderful contribution in the characterisation of *Pf*GluPho and Tim Bostick for helping in proofreading my manuscripts. To my fellow Ph.D colleagues K Buchholz, N Hiller, A Röseler, E Jortzik, S Koncarevic, K Sebastian thank you all. To all the other members of the group Doris, Antje and Raphael I appreciate the help I received from you in the course of this study.

It would have been an uphill task to carry out this research without the assistance of the Deutscher Akademischer Austausch Dienst (DAAD) who helped finance not only my stay in Germany throughout my study period but also the four months language course in Marburg.

Above all I am greatly indebted to my parents Mr. and Mrs. D. M. Musyimi and all my brothers, sisters and their families for their sacrifices, encouragement, patience, support and understanding during the course of this study. I extend my heartfelt and sincere thanks to Dr. P.L. M. Githua and Dr. M. N. Mwangome for their relentless support and encouragement throughout the course of this study.

Finally my heart felt gratitude to my wife Grace C. Mwangome, my daughter Lynn Ndimu Mwongela and my son Liam Musyimi Jnr III Mwongela for their sacrifices, patience, understanding and encouragement throughout this study.

List of Publications

Buchholz K, **Mailu B M**, Schirmer R H, Becker K. (2007). Structure based drug development against malaria. *Frontiers in Drug Design and Discovery*, 3: 225-255.

Rahlfs S, Koncarevic S, Iozef R, **Mailu B M**, Savvides S N, Schirmer R H, Becker K. (2008). Myristoylated adenylate kinase 2 of *Plasmodium falciparum* forms a heterodimer with myristoyl transferase. *Mol. Biochem. Parasitol.* **In press**.

Sturm N, **Mailu B M**, Jortzik E, Koncarevic S, Deponte S, Rahlfs S, Forchhammer K, Becker K. (2008). Identification of proteins targeted by the thioredoxin superfamily in *Plasmodium falciparum*. *PLoS Pathogens* **Submitted**.

Austin C J D, **Mailu B M**, Maghzal G J, Sanchez-Perez A, Rahlfs S, Zocher K, Arthur J, Becker K, Stocker R, Hunt N H, Ball H J. (2008). Recombinant mouse Indoleamine 2,3-dioxygenase like protein (rmIDO-2) utilise cytochrome b5 for optimal activity. *Biochemistry J.* **Submitted**.

Mailu B M, Rahlfs S, Becker K. (2008). Heterologous overexpression and characterisation of glucose 6-phosphate dehydrogenase-6-phosphogluconolactonase from *Plasmodium falciparum*. **In preparation**.

Conferences and Scientific Meetings

Mailu B M, Rahlfs S, Austin C J D, Hunt N H, Becker K. (2006). Further optimisation of the heterologous overexpression of mouse indoleamine 2, 3-dioxygenase. *22 Jahrestagung der Deutschen Gesellschaft für Parasitologie*, Wien, February 2006.

Mailu B M, Rahlfs S, Austin C J D, Hunt N H, Becker K. (2006). Optimisation of the heterologous overexpression of mouse indoleamine 2, 3-dioxygenase. *Second joint Ph.D Students Meeting of the Collaborative Research Centres SFB 544 Heidelberg and SFB 630 Würzburg*, Heidelberg, November 2006.

Austin C J D, **Mailu B M**, Sanchez-Perez A, McQuillan J, Astelbauer F, Rahlfs S, Aurther J, Becker K, Ball H J, Hunt N H. (2007). Indoleamine 2, 3-dioxygenase like protein 1 (INDOL 1). A novel heme containing tryptophan catabolising enzyme. *IUPAC Conference*, Turin Italy, July 2007.

Mailu B M, Hiller N, Fritz-Wolf K, Rahlfs S, Becker K. (2007). Interference with redox active proteins as a basis for the design of antimalarial drugs. *Drug Development Seminar*, Tübingen, April 2007.

Mailu B M, Rahlfs S, Becker K. (2008). Glucose 6-phosphate dehydrogenase-6-phosphogluconolactonase from *Plasmodium falciparum*. *Jahrestagung der Deutschen Gesellschaft für Parasitologie*, Hamburg, March 2008.

Summary

Plasmodium parasites are developing unacceptable levels of resistance to one drug after another and many insecticides are no longer useful against mosquitoes transmitting the disease. Years of vaccine research have produced few hopeful candidates and although scientists are redoubling the search, an effective vaccine is at best years away. Therefore there is need for identification of new drug targets and alternative antimalarial regimes. In response to this dire situation the study aimed at evaluating the pentose phosphate pathway of the malaria parasite *P. falciparum* in particular the bifunctional enzyme glucose-6-phosphate dehydrogenase-6-phosphogluconolactonase, understanding the kynurenine pathway of tryptophan metabolism in particular the enzymes indoleamine 2,3-dioxygenase (1 and 2) and unravelling more knowledge about the thioredoxin system networks in search for a new potential drug target and new drug alternatives.

The first two steps of the pentose phosphate pathway in *Plasmodium falciparum* are catalysed by the enzyme glucose 6-phosphate dehydrogenase-6-phosphogluconolactonase (*PfGluPho*) which is a unique bifunctional enzyme exclusively found in the genus *Plasmodium*. In spite of the importance of the role this enzyme plays in the parasite's pentose phosphate pathway as well as in overcoming oxidative stress, the characteristics of *PfGluPho* are still a mystery. For the first time *PfGluPho* has been successfully cloned, heterologously overexpressed and purified to homogeneity. The recombinant enzyme was found to be a hexamer which exhibits lower K_m values that favour substrate turnover by the parasite enzyme when compared to the human homologue. The steady state kinetics of *PfGluPho*'s glucose-6-phosphate dehydrogenase (*PfGluPho*'s G6PD) demonstrates that the enzyme follows an ordered sequential mechanism with NADP⁺ being the leading substrate. Three novel inhibitors of *PfGluPho*'s G6PD which are active at the lower micromolar range were identified and found to be non-competitive with respect to glucose-6-phosphate and NADP⁺. The study offers the first clear documentation of the cloning, heterologous overexpression, biochemical as well as kinetic characterisation, crystallisation and the first novel inhibitors of *PfGluPho*.

For 30 years, the established dogma regarding tryptophan catabolism was that the first step of the kynurenine pathway, the cleavage of the 2,3-double bond of the indole ring of tryptophan was performed by two enzymes, indoleamine 2,3-dioxygenase-1 (IDO-1) and tryptophan 2,3-dioxygenase (TDO). Recently, indoleamine 2,3-dioxygenase-2 (IDO-2) a third enzyme capable of performing this reaction has been discovered. Reported here is a study of the kinetic activity, pH stability, oligomeric structure as well as secondary structural features of recombinant mouse IDO-2 in direct comparison with mouse IDO-1. A screen for new more potent inhibitors of IDO-1 which lack the indole core and avoid the liability arising from the use of indole derivatives which have been reported to be neuroactive gave rise to compound 55D11 (K_i 0.05 μ M) which is more potent than the already existing IDO inhibitors. A structure activity study was done using various derivatives of compound 55D11 to determine

the elements that could be modified to increase potency. The study clearly demonstrates that IDO-1 and IDO-2 differ significantly in terms of their affinity for substrates as well as structure.

The malarial parasite *Plasmodium falciparum* possesses a functional glutathione and thioredoxin system comprising the redox-active proteins thioredoxin (Trx), glutaredoxin (Grx), and plasmoredoxin (Plrx) which all belong to the thioredoxin superfamily and share the active site motif Cys-X-X-Cys. A better understanding of the role of these members of the thioredoxin superfamily in *P. falciparum* as well as other systems could be achieved if more was known about their target proteins. Using thioredoxin affinity chromatography prepared by immobilising mutants of the redoxins lacking the resolving cysteine at the active site on CNBr activated sepharose, target proteins of *P. falciparum* cell extract were trapped. The covalently linked proteins were eluted with dithiothreitol and analyzed by matrix assisted laser desorption ionization time of flight (MALDI-TOF). Twenty one potential targets were identified for plasmoredoxin. Besides confirming known interacting proteins, potential target candidates involved in processes such as; protein biosynthesis, energy metabolism and signal transduction were identified. Further confirmations of the interaction of plasmoredoxin and the target proteins were done using BIAcore surface plasmon resonance experiments.

Zusammenfassung

Der Malaria-Parasit *Plasmodium* entwickelt bemerkenswert hohe Resistenzen gegenüber einem Medikament nach dem anderen. Außerdem verlieren viele Insektizide, die gegen die Überträger-Moskitos eingesetzt werden, an Wirkung. Jahrelange Forschung an Impfstoffen gegen Malaria hat bisher nur wenige hoffnungsvolle Kandidaten erbracht und obwohl Wissenschaftler Ihre Bemühungen verstärken, ist eine effektive Impfung bestenfalls immer noch Jahre entfernt. Deshalb sind dringend neue Zielmoleküle für die Medikamentenentwicklung zu identifizieren, die zu alternativen Behandlungsmethoden führen können. Diese Situation vor Augen, waren es Ziele dieser Arbeit, i) das bifunktionelle Enzym Glukose-6-Phosphatdehydrogenase-6-Phosphogluconolactonase aus dem Pentosephosphatweg des Parasiten als potentielles Wirkungsziel zu bestätigen, ii) den Kynurenin-Stoffwechselweg, insbesondere die Enzyme Indolamin 2,3-Dioxigenase (1 und 2) der Maus als Modell näher zu charakterisieren und iii) mehr über das Redox-Netzwerk des Malaria-parasiten zu erfahren, um neue mögliche Zielmoleküle aufzuzeigen.

Die ersten zwei Schritte des Pentosephosphatweges werden in *Plasmodium falciparum* durch das Enzym Glukose-6-Phosphatdehydrogenase-6-Phosphogluconolactonase (*Pf*GluPho) katalysiert. Dieses ist ein einzigartiges bifunktionelles Enzym, das bisher nur in *Plasmodien* gefunden wurde. Obwohl diesem Enzym im Pentosephosphatweg des Parasiten und damit auch in der Bekämpfung oxidativen Stresses eine enorme Bedeutung zukommt, ist das Enzym aus *P. falciparum* nicht besonders gut charakterisiert, da es bisher nicht kloniert werden konnte. Zum ersten Mal konnte *Pf*GluPho jetzt kloniert und überexprimiert werden und das Genprodukt konnte bis zur Reinheit gebracht werden. Es wurde gezeigt, dass das rekombinante Enzym als Hexamer vorliegt, welches niedrigere Km-Werte im Vergleich zu seinen humanen Orthologen aufweist, die einen bevorzugten Substratumsatz durch das parasitäre Enzym aufzeigen. Kinetische Untersuchungen zeigen, dass der Glukose-6-Phosphatdehydrogenase (G6PD)-Teil von *Pf*GluPho einem geordneten Mechanismus folgt, bei dem NADP⁺ das erste Substrat ist. Drei neue Inhibitoren, die den G6PD-Teil des Enzyms im unteren mikromolaren Konzentrationsbereich hemmen, konnten gefunden werden und zeigten sich gegenüber Glukose-6-Phosphat und NADP⁺ als nicht-kompetitiv. Somit zeigt diese Arbeit die Klonierung, heterologe Expression, biochemische und kinetische Charakterisierung von *Pf*GluPho auf, sowie darüberhinaus die Kristallisation und erste, neue Inhibitoren.

Seit 30 Jahren ist es ein etabliertes Dogma, dass im Tryptophan-Katabolismus der erste Schritt des Kynurenin-Stoffwechselweges die 2,3-Doppelbindung des Indolringes des Tryptophans durch zwei Enzyme gespalten werden kann: einerseits durch Indolamin 2,3-Dioxigenase (IDO), anderseits durch Tryptophan 2,3-Dioxigenase (TDO). Kürzlich konnte mit IDO-2 ein drittes Enzym in der Maus und im Menschen entdeckt werden, das in der Lage ist, diese Reaktion zu vollziehen. In dieser Arbeit sind Daten zur Kinetik, pH-Stabilität, zu oligomeren Strukturen, sowie Besonderheiten der Sekundärstruktur von rekombinanter IDO-2

der Maus im direkten Vergleich mit rekombinanter IDO-1 der Maus erhoben worden. Die Suche nach neuen, effektiveren Inhibitoren für IDO-1, die den Indolkern nicht mehr besitzen und somit weniger neurotoxisch sein könnten, führte zu 55D11 (K_i : 0,05 μM), einer Substanz aus einer öffentlichen Sammlung von Naturstoffen. Zahlreiche Derivate von 55D11 wurden untersucht, um Molekülreste zu zeigen, die verändert werden können und die Aktivität noch erhöhen. Die Untersuchungen zeigen, dass sich IDO-1 und IDO-2 eindeutig hinsichtlich ihrer Affinität zu Substraten, aber auch in ihrer Struktur unterscheiden.

Der Malaria-Parasit *Plasmodium falciparum* besitzt ein funktionelles Glutathion-, sowie ein Thioredoxin-System, die unter anderem aus den redox-aktiven Proteinen Thioredoxin (Trx), Glutaredoxin (Grx) und Plasmoredoxin (Plrx) bestehen, die alle zur sogenannten Thioredoxin-Superfamilie gehören und das Motiv Cys-X-X-Cys im aktiven Zentrum besitzen. Wenn mehr Interaktionspartner dieser Redoxproteine bekannt wären, könnte die jeweilige Rolle der Redoxine in ihrem Netzwerk besser verstanden werden. Deshalb wurden Thioredoxin-Affinitätschromatographien an CNBr-aktivierter Sepharose durchgeführt. Hierzu wurden immobilisierte Mutanten der Redoxine, denen das sogenannte „*resolving cysteine*“ aus ihrem aktiven Zentrum fehlt, mit Zellextrakt aus *P. falciparum* versetzt. Kovalent an die Fängerproteine (Redoxine) gebundene Reaktanden wurden mit Dithiothreitol eluiert und mittels (Maldi-ToF) analysiert. Einundzwanzig potentielle Zielproteine wurden für Plasmoredoxin als mögliche Interaktionspartner identifiziert. Neben bekannten Interaktionspartnern waren darunter auch Kandidaten, die eine mögliche Redoxregulierung der Proteinbiosynthese, des Energiemetabolismus sowie der Signaltransduktion in *Plasmodium* vermuten lassen können. Weitere Untersuchungen, um diese Wechselwirkungen zu bestätigen, wurden mit Plasmoredoxin und einigen seiner potentiellen Interaktionspartner mithilfe von BIACORE Oberflächen-Resonanzexperimenten durchgeführt.

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